An introduction to phylogenetic networks

Steven Kelk

Department of Knowledge Engineering (DKE) Maastricht University

Email: steven.kelk@maastrichtuniversity.nl Web: http://skelk.sdf-eu.org





Genome sequence, comparative analysis and haplotype structure of the domestic dog

> Lindblad-Toh et al, Nature 2005



(Almost) everything begins with Multiple Sequence Alignment

	* *	
Q5E940_BOVIN	MPREDRATWKSNYFLKIIQLLDDYPKCFIVGADNVGSKQMQQIRMSLRGK-AVVLMGKNTMMRKAIRGHLENNPALE	76
RLA0 HUMAN	MPREDRATWKSNYFLKIIQLLDDYPKCFIYGADNYG <mark>S</mark> KOMQQIRMSLRGK-AYYLMGKNTMMRKAIRGHLENNPALE	76
RLA0 MOUSE	MPREDRATWKSNYFLKIIQLLDDYPKCFIYGADNYGSKQMQQIRMSLRGK-AVYLMGKNTMMRKAIRGHLENNPALE	76
RLAO_RAT	MPREDRATWKSNYFLKIIQLLDDYPKCFIYGADNYG <mark>S</mark> KQMQQIRMSLRGK-AYYLMGKNTMMRKAIRGHLENNPALE	76
RLA0 CHICK	MPREDRATWKSNYFMKIIQLLDDYPKCFVVGADNVGSKQMQQIRMSLRGK-AVVLMGKNTMMRKAIRGHLENNPALE	76
RLA0 RANSY	·MPREDRATWKSNYFLKIIQLLDDYPKCFIYGADNYGSKQMQQIRMSLRGK-AVYLMGKNTMMRKAIRGHLENNSALE	76
Q7ZUG3 BRARE	MPREDRATWKSNYFLKIIQLLDDYPKCFIYGADNYGSKQMQTIRLSLRGK-AVYLMGKNTMMRKAIRGHLENNPALE	76
RLA0 ICTPU	MPREDRATWKSNYFLKIIQLLNDYPKCFIYGADNYGSKQMQTIRLSLRGK-AIVLMGKNTMMRKAIRGHLENNPALE	76
RLA0 DROME	MVRENKAAWKAQYFIKVVELFDEF <mark>P</mark> KCFIVGADNVG <mark>S</mark> KQMQNIRTSLRGL-AVVLMGKNTMMRKAIRGHLENNPQLE	76
RLA0 DICDI	MSGAG-SKRKKLFIEKATKLFTTYDKMIVAEADFVG <mark>S</mark> SQLQKIRKSIRGI-GAVLMGKKTMIRKVIRDLADSKPELD	75
Q54LP0 DICDI	MSGAG-SKRKNVFIEKATKLFTTYDKMIVAEADFVG <mark>S</mark> SQLQKIRKSIRGI-GAVLMGKK <mark>T</mark> MIRKVIRDLADSK <mark>P</mark> ELD	75
RLA0 PLAF8	MAKLSKQQKKQMYIEKLSSLIQQYSKILIVHVDNVGSNQMASVRKSLRGK-ATILMGKNTRIRTALKKNLQAVPQIE	76
RLA0_SULAC	MI <mark>G</mark> LAYTTTKK IAKWK <mark>YDEYAELTEKLKTHKTIIIAN IEGFP</mark> ADKLHEIRKKLRGK-ADIKYTKNNLFN IALKNAGYDTK	79
RLA0 SULTO	MRIMAVITQERKIAKWKIEEVKELEQKLREYHTIIIANIEGFPADKLHDIRKKMRGM-AEIKVTKNTLFGIAAKNAGLDVS	80
RLA0 SULSO	MKRLALALKQRKVASW <mark>K</mark> LEEVKELT <mark>E</mark> LIKNSNTILI <mark>G</mark> NL <mark>EGFP</mark> ADKLHEIRKKLRGK-A <mark>TIKVTKNT</mark> LFKIAAKNAGIDIE	80
RLA0 AERPE	MSVVSLVGQMYKREK <mark>PIPEWK</mark> TLMLRELE <mark>E</mark> LFSKHRVVLFADLTG TPT FVV <mark>G</mark> RVRKKLWKK-YPMMVAKKRIILRAMKAAGLELDDN	86
RLA0 PYRAE	-MMLAI <mark>G</mark> KRRYARTRQ <mark>YP</mark> AR <mark>K</mark> AKIASEAT <mark>E</mark> LLQK <mark>YP</mark> YAFLFDLH <mark>G</mark> LS <mark>S</mark> RILHE <mark>YR</mark> YRL <mark>R</mark> RY-GAIKIIKPTLFKIAFTKAYGGIPAE	85
RLA0 METAC	MAEERHHTEHIPQWKKDEIENIKELIQSHKYFGMYGIEGILATKMQKIRRDLKDY-AYLKYSRNTLTERALNQLGETIP	78
RLA0 METMA	MAEERHHTEHIPQWKKDEIENIKELIQSHKYFGMYRIEGILATKIQKIRRDLKDY-AYLKYSRNTLTERALNQLGESIP	78
RLA0 ARCFU	MAAVRCS <mark>PPEYK</mark> VRAVEEIKRMISSK <mark>P</mark> VVAIVSFRNVPACOMOKIRREFRGK-AEIKVVKNTLLERALDALCCDYL	75
RLA0_METKA	MAYKAK <mark>G</mark> Q <mark>PP</mark> SGYEPKVAEWK <mark>RREVKELKELMDEYENVGLVDLEGIPAPQLQEIR</mark> AKL <mark>R</mark> ERD <mark>TIIRMSRNT</mark> LMRIALEEKLDERPELE	88
RLA0_METTH	MAHVAEWKKKEVQELHDLIK <mark>GY</mark> EVVGIANLADIPAR <mark>Q</mark> LQKMRQTLRDS-ALIRMSKKTLISLALEKAGRELENVD	74
RLAO METTL	MITAESEHKIAPWKIEEVNKLKELLKNGQIVALVDMMEVPARQLQEIRDKIR-GTMTLKMSRNTLIERAIKEVAEETGNPEFA	82
RLA0 METVA	. ––––––MIDAKSEHKIA <mark>P</mark> WKIEEVNALKELLKSANVIALIDMMEV <mark>P</mark> AV <mark>Q</mark> LQEIRDKIR–DQMTLKMSRNTLIKRAVEEVAEETGNPEFA	82
RLA0_METJA	. –––––––METKYKAHYA <mark>P</mark> WK <mark>IEEYKTLKGLIKSKPYYAIYDMMDYPAPQLQEIR</mark> DKI <mark>R</mark> –DKYKL <mark>RMSRNT</mark> LII <mark>RALKEAAE</mark> EINN <mark>P</mark> KLA	81
RLA0_PYRAB	MAHVAEWKKKEVEELANLIKS <mark>YP</mark> VIALVDVSSMPAY <mark>P</mark> LSQMRRLIREN <mark>GGLLRVSRNT</mark> LIELAIKKAAQEL <mark>GKP</mark> ELE	77
RLA0_PYRHO	MAHVAEWKKKEVEELAKLIKS <mark>YP</mark> VIALVDVSSMPAY <mark>P</mark> LSQMRRLIREN <mark>GGLLRVSRNT</mark> LIELAIKKAAKEL <mark>GKP</mark> ELE	77
RLA0_PYRFU	MAHVAEWKKKEVEELANLIKS <mark>YP</mark> VVALVDVSSMPAY <mark>P</mark> LSQMRRLIRENNGLLRV <mark>SRNT</mark> LIELAIKKVAQELGKPELE	77
RLA0_PYRKO	MAHVAEWKKKEVEELANIIKS <mark>YP</mark> VIALVDVAGVPAY <mark>P</mark> LSKMRDKLR-GKALLRVSRNTLIELAIKRAAQELGOPELE	76
RLA0_HALMA	MSAESERKTET IPEWKQEEVDAIVEMIESYESVGVVNIAGIPSRQLQDMRRDLHGT - AELRVSRNTLLERALDDVDDGLE	79
RLA0_HALVO	MSESEVRQTEVIPQWKREEVDELVDFIESYESVGVVGVAGIPSRQLQSMRRELHGS-AAVRMSRNTLVNRALDEVNDGFE	79
RLA0_HALSA	MSAEEQRTTEEV <mark>P</mark> EWK <mark>RQEVAELVDLLETY</mark> DSVGVVNVTG <mark>IPS</mark> KQLQDMRRGLHGQ-AALRMSRNTLLVRALEEAGDGLD	79
RLA0_THEAC	MKEVSQQ <mark>K</mark> KELVNEITQRIKASRSVAIVD <mark>I</mark> AGIRTRQIQDIRGKN <mark>RG</mark> K-INLKVIKKTLLFKALENLGDEKLS	72
RLA0_THEVO	MRKIN <mark>P</mark> KKKEIVSELAQDITKSKAVAIVDIKGVRIRQMQDIRAKNRDK-VKIKVVKKTLLFKALDSINDEKLT	72
RLA0_PICTO	MTE <mark>PAQWK</mark> IDFVKNLENEINSRKVAAIVSIK <mark>G</mark> LRNN <mark>EFQ</mark> KI <mark>R</mark> NSIRDK-ARIKV <mark>SR</mark> ARLLRLAIEN <mark>TG</mark> KNNIV	72
ruler	$1 \dots \dots 10 \dots \dots 20 \dots \dots 30 \dots \dots 40 \dots \dots 50 \dots \dots 60 \dots \dots 70 \dots \dots 80 \dots \dots 90$	

Dominant methods for building phylogenetic trees

- Character-based methods
 - Maximum Parsimony (MP)
 - Maximum Likelihood (ML)
- Bayesian methods (Markov Chain Monte Carlo MCMC)
- Distance-based methods
 - Neighbour Joining
 - UPGMA
- "Supertree" methods: glueing together smaller subtrees



Sequence 1	Т	G	С
Sequence 2	Т	А	С
Sequence 3	А	G	G
Sequence 4	А	А	G





From: http://artedi.ebc.uu.se/course/X3-2004/Phylogeny/Exercises/mp.html

Homology of HXT1p to other fungal hexose transporters.





Voegele R T et al. PNAS 2001;98:8133-8138 The role of haustoria in sugar supply during infection of broad bean by the rust fungus *Uromycesfabae*





There is more to life than trees

• All these methods assume that a (single) tree is the best way to model the underlying evolution.

• If this is not true, then we have a problem, because there is a high risk that the output of tree-building algorithms will then be meaningless.

- Sometimes there are clues about this:
 - Algorithms build very badly supported trees
 - Extra knowledge about the underlying evolutionary mechanisms
- But in general it is **dangerously easy** to confuse non-treelike evolution with a **noisy tree signal**.
- Therefore critical to understand and model underlying mechanisms.

Why might we get weak support for a tree?

"Noisy tree"

Data *does* fit a single tree, weak support is only a consequence of "noise"

"Trees in trees"

Data consists of multiple different tree signals...but both gene and species evolution are still ultimately treelike (e.g. due to incomplete lineage sorting, gene loss, gene duplication)

"Trees in networks"

Data consists of multiple different tree signals...gene evolution is treelike, but species evolution is no longer treelike (e.g. hybridization, horizontal gene transfer)

"Reticulation"

Inherently nontreelike (reticulate) phenomena, such as meiotic, sexual recombination







Data-display networks (1)



From: Daniel Huson, ISMB-Tutorial 2007: Introduction to Phylogenetic Networks



Data-display networks (2)



A phylogenetic network. The network was generated by Neighbor-Net for a sequence-based data set comprising of Salmonella isolates that originally appeared in [17]. A detailed network-based analysis of this data is presented in [2], where the strains indicated in bold-face are tested for the presence of recombination. Note that the network is planar (that is, it can be drawn in the plane without any crossing edges), and that parallel edges in the network represent bipartitions of the data.

Bryant et al. Algorithms for Molecular Biology 2007 2:8 doi:10.1186/1748-7188-2-8

Sty19*



Data-display networks (3)

• Data-display networks do not automatically generate a hypothesis of *what actually happened.*

• They restrict themselves to showing how and where the input data is not tree-like.

• Some biologists are starting to use these networks, to perform what David Morrison calls "Exploratory Data Analysis (EDA)".

• For an experienced biologist, looking to apply his/her own expert knowledge to explain what actually happened (i.e. ad-hoc hypothesis generation), such a tool can give very important insights.





Evolutionary phylogenetic networks

- Used to explicitly model reticulate evolution:
 - Hybridization
 - Horizontal Gene Transfer (HGT)
 - Recombination
- Reticulation events have an explicit biological interpretation
- Usually rooted, with an explicit "direction" of evolution
- Underlying mathematical abstractions are often similar, despite different scale levels of interpretation





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Different models and scales, but always rooted, directed acyclic graphs (DAGs)



Ancestral Recombination Graph (ARG)



(f) nad7

Horizontal Gene Transfer (HGT)



"Softwired cluster" network



Constructing evolutionary phylogenetic *networks*

• It's important to ask ourselves several questions:

1. **MODEL**: What are we trying to **model** exactly? Is it biologically realistic?

2. OBJECTIVE: What do we consider to be an "optimal" solution within that model?

3. TRACTABILITY: Is there any hope of developing efficient algorithms to compute optimal solutions?

• Extremely challenging to simultaneously answer these questions well!

• In the meantime: many different models, algorithms, packages

Several case studies

- 1. A "direct" method : constructing Ancestral Recombination Graphs (ARGs) by modelling crossover events.
- 2. "The trees within" : methods which analyse phylogenetic networks based on the set of trees **contained within them**.
 - a) Extensions to Maximum Parsimony (MP) and Maximum Likelihood (ML)
 - b) Parsimoniously embedding gene trees in species networks



Case study 1: constructing Ancestral Recombination Graphs (ARGs)





- Reticulations represent chromosomal crossover (mostly single crossover, sometimes multiple crossover). Sometimes also gene conversion.
- Mutation model is the "infinite sites" model: at most one mutation per site (0 to 1, or 1 to 0).

• Goal is to construct an ARG with a **minimum number** of reticulation events.



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Case study 1: constructing Ancestral Recombination Graphs (ARGs)

- Programs for constructing ARGs include HAPBOUND, SHRUB, BEAGLE
- Extensive interest and research from the theoretical computer science community (e.g. Dan Gusfield)
- Issues:
 - Difficult to solve (NP-hard, also difficult in practice)
 - Modelling of homoplasy (recurrent and back mutation) is in its infancy (infinite sites model excludes this)
 - Rigid biological model (crossover)
 - Software implementations still rather experimental
 - Standard phylogenetic concepts such as bootstrapping, branch-lengths etc. are not considered







Case study 2(a): extensions to Maximum Parsimony and Maximum Likelihood

- The group of Luay Nakhleh (Rice University, USA) is very interested in this.
- The general idea is to define the parsimony/likelihood score of a network, as a function of the set of trees contained within it.
- Software: PHYLONET, NEPAL
- Issues:
 - Again, a very specific (and thus rigid) model
 - Assumed independence of characters leads to problems
 - More reticulations = better score, so when do we stop adding reticulations?
 - Even "small" variant (e.g. here is a network, compute the best parsimony score for it) is algorithmically challenging

• Algorithms for the "big" variant (i.e. find me the best network) are still very basic



From: Jin, G., Nakhleh, L., Snir, S., Tuller, T.: *Inferring phylogenetic networks by the maximum parsimony criterion: A case study.* Molecular Biology and Evolution 24(1), 324–337 (2007).

MP analysis based on the ribosomal protein gene *rps11* of a group of 47 flowering plants, which was analysed by Bergthorsson et al (2003)



Case study 2(b): combining multiple gene trees into a single species network



• Recall this example:



Four gene trees contained in the species network

- Input: a set of gene trees
- Output: a species network that contains all the input gene trees and which has a minimum number of reticulations

From: Fast computation of minimum hybridization networks, Benjamin Albrecht, Celine Scornavacca, Alberto Cenci and Daniel H. Huson, to appear in Bioinformatics (2011). (a) (b) T monococcum T monococcum T urartu T urartu • Ae tauschii Ae tauschii Ae comosa Ae_comosa Ae uniaristata Ae uniaristata Ae bicornis Ae bicornis Ae_longissima Ae longissima Ae sharonensis Ae sharonensis Ae_speltoides Ae speltoides Hordeum Hordeum

Fig. 3. The two consensus trees computed from 100 bootstrap replicates for the matK (**a**) and PinA (**b**) datasets.





Fig. 4. The three hybridization networks obtained by the described algorithm for the matK and PinA consensus trees of Figure 3.



So...how far have we come? What do we still have to do?

Summary of progress/problems



• Data-display networks are starting to attract attention from the biological community as an instrument for Exploratory Data Analysis. But still very marginal. The software is there, however, and in time they will I think become mainstream tools.

• Evolutionary phylogenetic networks – those which try and hypothesise *what* actually happened – have the potential to become a very powerful tool for biologists. But at the moment they are, in practice, hardly used at all:

•(Severe) computational intractability.

 Algorithms in general do not generate multiple optimal solutions and have no network equivalent of common "tree" concepts such as bootstrapping, branch-lengths etc.

• Very many biological phenomena can cause phylogenetic signals to be nontreelike. At the moment there is **no consensus amongst biologists** how to model these.

Ideas for the future (1/3)

Remember the context...

•"Everyone" seems to build phylogenetic trees, but "nobody" uses software for (evolutionary) phylogenetic networks. What's going wrong?

• Remember that the concept of "phylogenetic network" covers a very wide array of disparate evolutionary phenomena, many of which are still poorly understood.

• Is it realistic, then, to expect that there is **one model/software package to rule them all?** Perhaps it can and should remain a specialised phenomenon, adapted ad-hoc on a case-by-case basis?

Ideas for the future (2/3)



• Ensure that the software gives the biologists what they want

• Phylogenetic tree construction is so standardized that certain concepts (such as bootstrapping: a measurement of solution robustness) are seen as essential.

• It's therefore important to develop (standardized?) equivalents for phylogenetic network construction; they are not yet there.

 There is some reason for optimism here, since the question "how confident are you that this is the right solution?" can at least partially be answered in a model-neutral way.



Ideas for the future (3/3)

Better co-ordination between computer scientists and biologists

• Scientists working on the algorithmic efficiency side of phylogenetic networks rarely have more than a superficial understanding of the biological model. Much more contact with biologists needed.

• *"The future of phylogenetic networks"* – modelling workshop at Lorentz Center in Leiden, October 2012.

Lorentz Center

International Center for workshops in the Sciences

Current Workshop	Overview	Back	Print	Home	Search	Contact
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The Future of Phylogenetic Networks from 15 Oct 2012 through 19 Oct 2012

Venue: Lorentz Center@Oort

- Description and aim of the workshop
- Registration form
- Participants
- Program
- Abstracts
- Event report
- Presentations
- Scientific organizers: Leo van Iersel (Amsterdam, Netherlands) Steven Kelk (Maastricht, Netherlands) David Morrison (Uppsala, Sweden) Leen Stougie (Amsterdam, Netherlands)



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Slides: http://www.lorentzcenter.nl/lc/web/2012/515/info.php3?wsid=515

(Slide by Leo van Iersel)

What do biologists want?

19 October 2012 The Future of Phylogenetic Networks

Lorentz Center Leiden, The Netherlands (Slide by Leo van Iersel)

What do biologists want?

- Biologists don't know what they want
- Depends on data and goals
- Changes all the time

Finally...further reading

• Luay Nakhleh, "Evolutionary phylogenetic networks: models and issues." In: The Problem Solving Handbook for Computational Biology and Bioinformatics, L. Heath and N. Ramakrishnan (editors). Springer, 125-158, 2010.

• Daniel Huson, Regula Rupp and Celine Scornavacca, *"Phylogenetic Networks"*, Cambridge University Press, 2010

• David Morrison, *"An introduction to phylogenetic networks",* RJR-productions, 2011

• *"The genealogical world of phylogenetic networks",* http://phylonetworks.blogspot.nl/



Concepts, Algorithms and Applications

CAMBRIDGE

Daniel H. Huson Regula Rupp Celine Scornavacca





Extra unused slides

Case study 2(b): combining multiple gene trees into a single species network

• There has been a huge amount of research from the theoretical computer science community for the case when the input consists of exactly two binary gene trees

- The result is a lot of very nice math, and increasingly fast algorithms (such as HYBRIDNET and an algorithm in DENDROSCOPE 4)
- Issues:
 - No software exists to reliably compute optimal solutions for three or more trees, even when binary
 - Multiple solutions? Branch lengths? Bootstrapping?
 - Rooting problems



- Rooted triplets: phylogenetic trees with only 3 leaves
- The idea is that it might be easier to build lots of very small trees (rooted triplets) and to merge them into a single network, then to try and construct the network in one go
- Rooted triplets can be inferred directly/ad-hoc or extracted from gene trees
- Idea is similar to trees i.e. combine them into a single network such that the number of reticulations is minimised



- For example. Suppose I want to reconstruct a plausible evolution for the species set {w,x,y,z}.
- I am given a set of rooted triplets zw|x, yx|w, xy|z, wz|y. (Note zw|x = wz|x.)





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• There are several programs for building networks from rooted triplets (LEVEL2, LEV1ATHAN, SIMPLISTIC)

• In theory the advantage for the user (above trees) is that it is not necessary to first construct entire gene trees; the user can instead choose to specify only high-quality fragments of them as input.

• Also possible to construct the rooted triplets from heterogeneous sources (because abstraction is "value free").

- Issues:
 - How do we generate good rooted triplets in the first place?
 - Input-side demands to ensure tractability are too restrictive
 - Small amount of noise can inflate the number of reticulations
 - Multiple solutions? Branch lengths? Bootstrapping?
 - Lack of memory: topology is not preserved



Figure 5. The level-1 network on the right with a single reticulation represents the union of the clusters (and triplets) obtained from the three trees on the left. However, any network that displays all three trees will have at least two reticulations and have level at least two.







From: Multigenic phylogeny and analysis of tree incongruences in Triticeae (Poaceae), Escobar et al, BMC Evolutionary Biology 2011, 11:181

Figure 3 Multigenic network of Triticeae. Network obtained from the 27 individual gene trees modified with PhySIC_IST [56] using a correction threshold of 0.9 (see details in Methods).



Fig. 16 A gene tree (solid lines) evolving within the branches of the species tree, where the gene tree topology is identical to that of T_2 in Fig. 1(b). The gene tree differs from the species tree due to (incomplete) lineage sorting.

From: L. Nakhleh, "Evolutionary phylogenetic networks: models and issues." In: The Problem Solving Handbook for Computational Biology and Bioinformatics, L. Heath and N. Ramakrishnan (editors). Springer, 125-158, 2010.