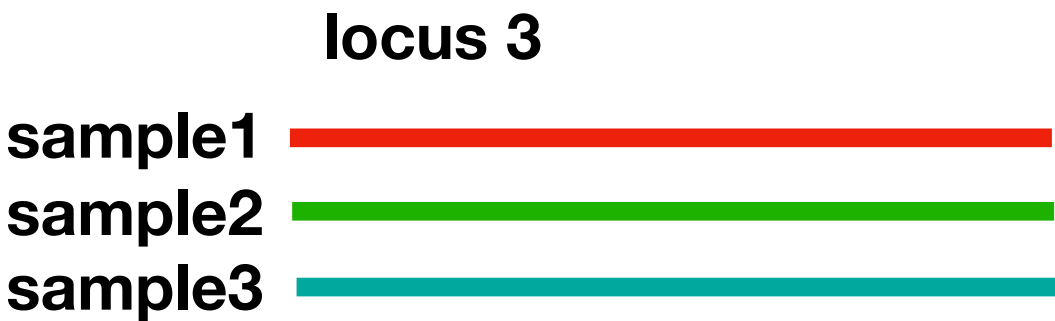
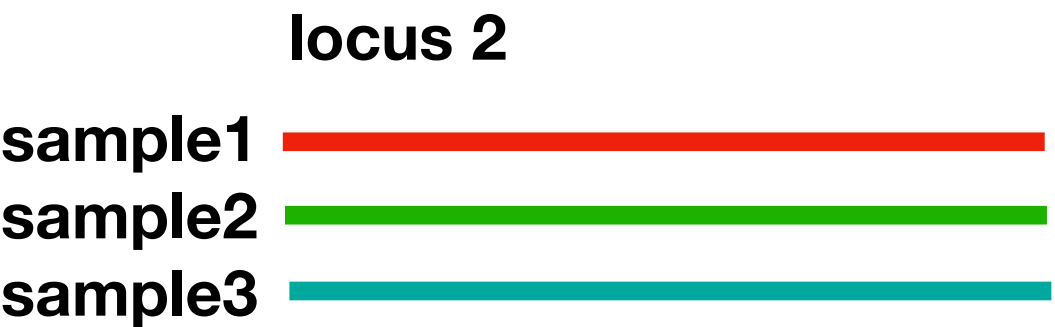
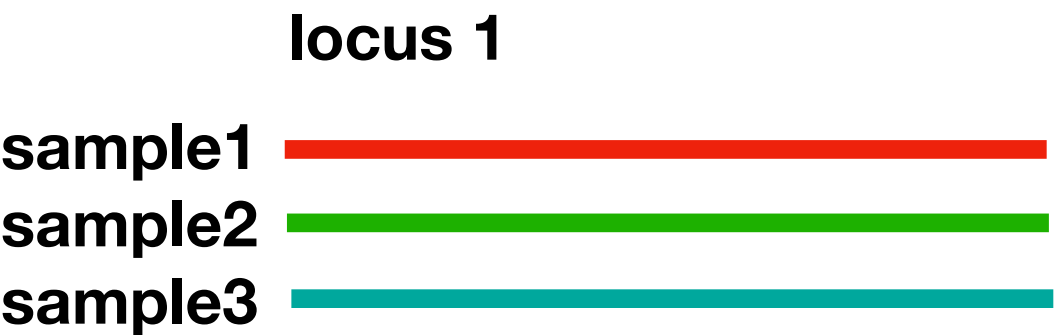


Post-processing of enrichment data

Reporter: Hao Yuan

Output of assembling pipeline

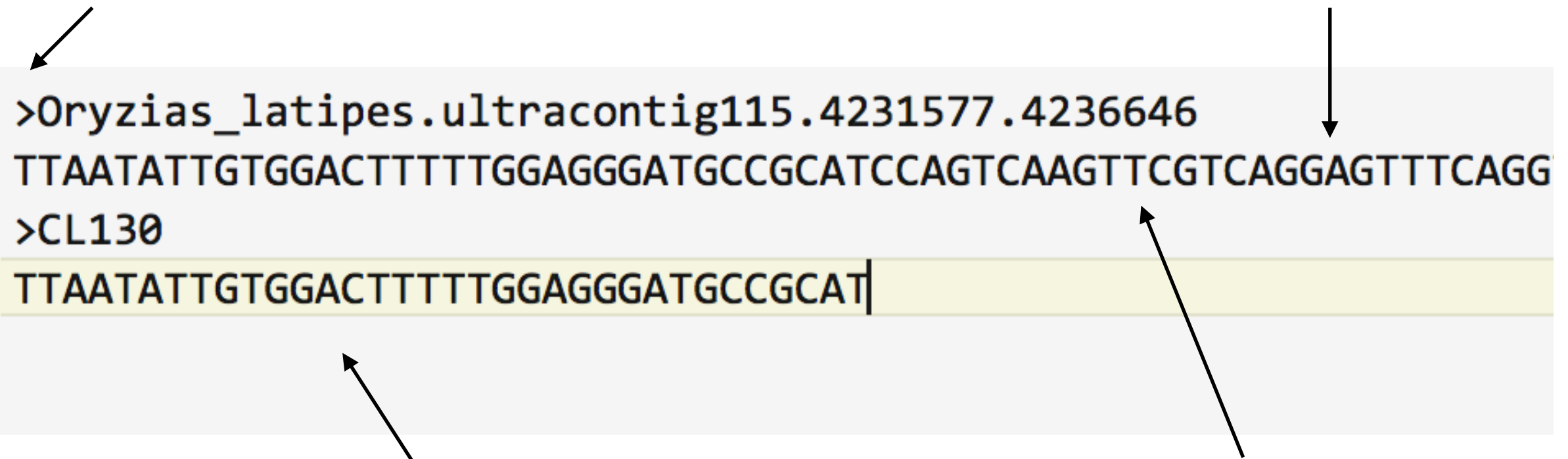


Sequences targeting the same loci

Output until here looks like this

A “>” before the sequence name

After name is sequence



```
>Oryzias_latipes.ultracontig115.4231577.4236646  
TTAATATTGTGGACTTTTTTGGAGGGATGCCGCATCCAGTCAAGTTCGTCAGGAGTTTCAGG  
>CL130  
TTAATATTGTGGACTTTTTTGGAGGGATGCCGCAT|
```

The diagram shows a FASTA format output. The first line is a header starting with a greater-than sign (>) followed by the sequence name. The second line is the sequence. The third line is another header starting with > followed by the sequence name. The fourth line is the sequence, which is highlighted with a yellow background and ends with a vertical bar. Arrows point from the text labels to the corresponding parts of the output: one arrow points to the > symbol in the first header, another points to the sequence line of the first header, a third points to the > symbol in the second header, and a fourth points to the sequence line of the second header.

First one is the sequence of reference

Following is the sequence of enriched sample

This called **fasta** format. File suffix is “**fa**”, “**fas**” or “**fasta**”

Assembled contigs are coding sequences

Start from the
first codon

End at from the
third codon

TAT**AAC****CTG**

Length of nucleotide can
be exactly divided by 3

Y **N** **L**

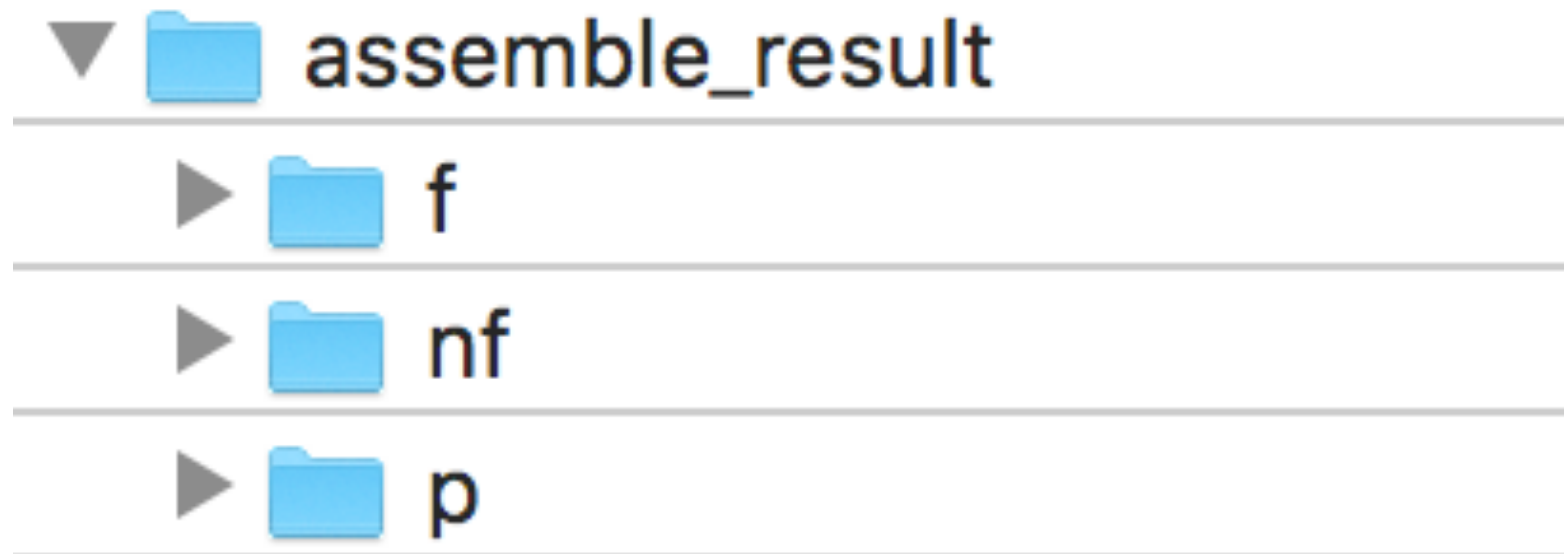
No stop codon in amino acid sequences

Output will be placed under "assemble_result" including 3 folders:

1) nf: folder containing full coding nucleotide sequences

2) f: folder containing coding sequences with flankings

3) p: folder containing amino acid sequences



Further processing

Data manipulation

Multiple sequences alignment

Filter poorly aligned regions

Filter for other purpose

Detect cross-sample contamination

Summary statistics

Data manipulation

Remove poorly enriched taxa

enriched_gene.txt

```
total    4435

Sample   Num. of enriched genes   Percentage of enriched genes(%)
sample1  3262      73.6
sample2  3253      77.3
sample3  3356      75.7
sample4  3516      79.3
sample5  410       9.24
```

“—deselected_taxa” option of [pick_taxa.pl](#)

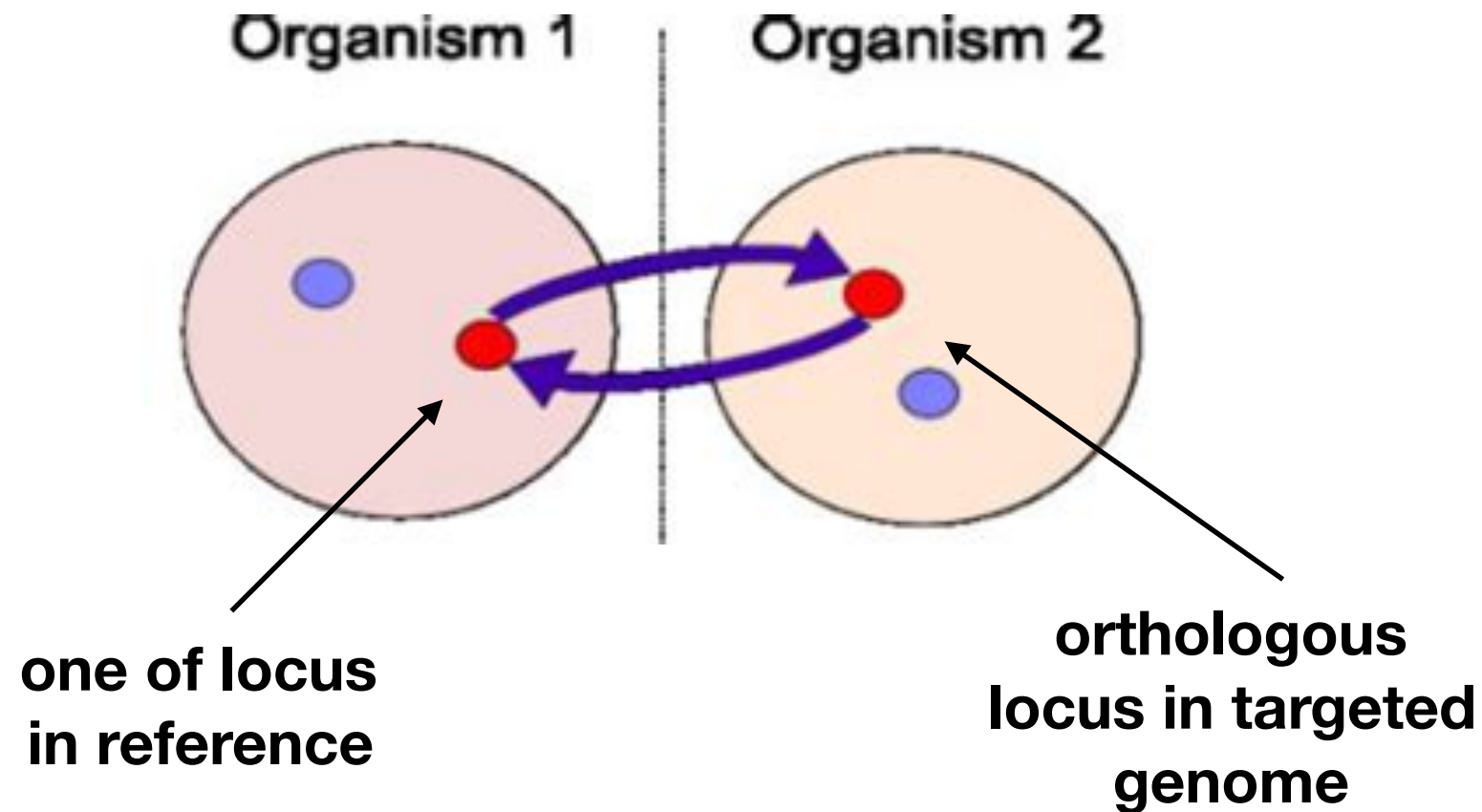
Get loci with certain data completeness level

“—min_seq” option of [pick_taxa.pl](#)

$$\begin{array}{ccc} & 10 * 0.8 = 8 & \\ \swarrow & & \nwarrow \\ \text{number of} & & \text{completeness} \\ \text{total sample} & & \text{level} \end{array}$$

Data manipulation

Extract loci from existing genomes



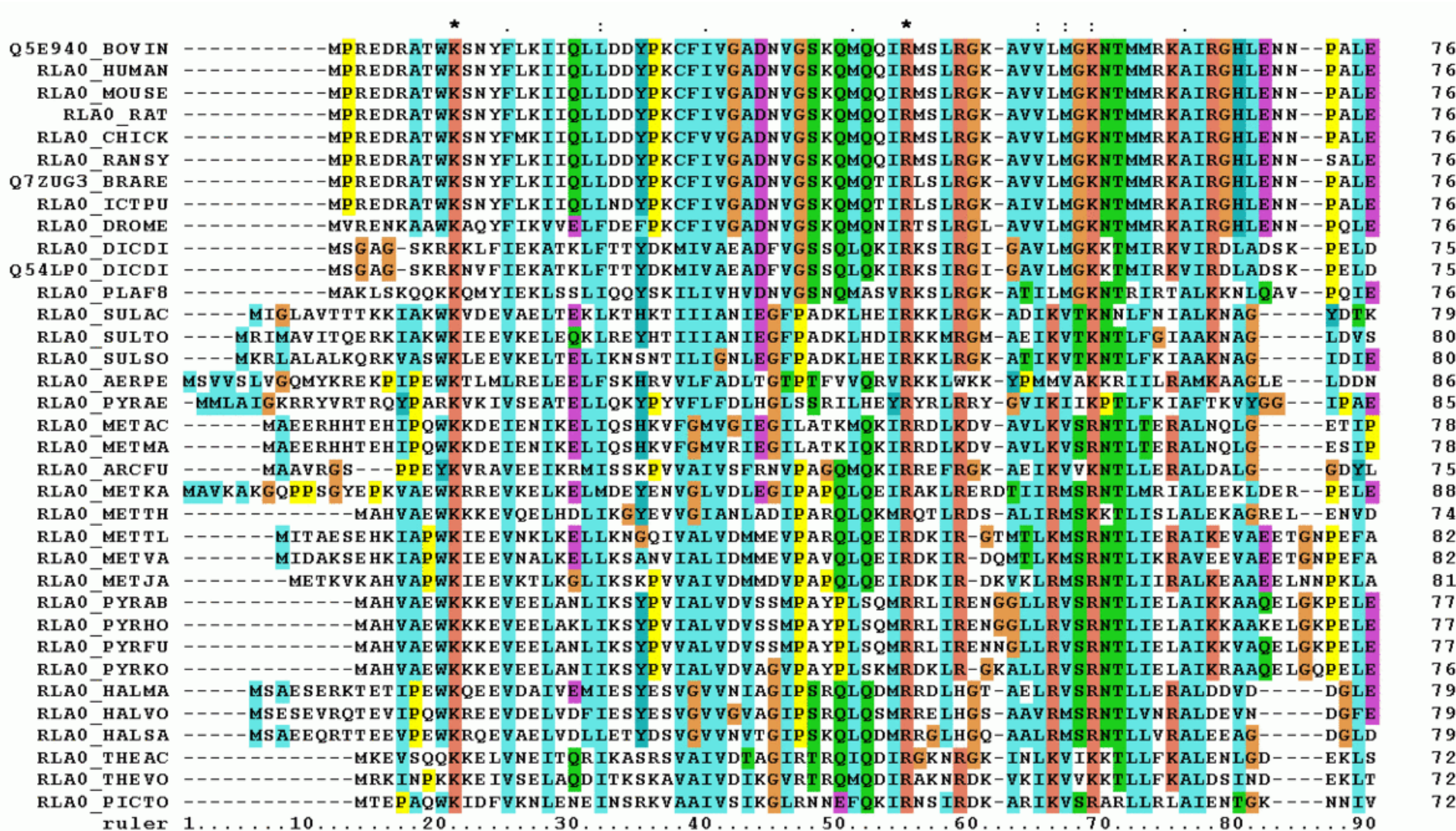
use `merge_loci.pl` to add extracted sequences to enriched dataset

`get_orthologues.pl`
`merge_loci.pl`

Multiple sequences alignment

specify “`–non_codon_aln`”
option of `mafft_aln.pl` if align
sequences with flanks

Reads → Contigs



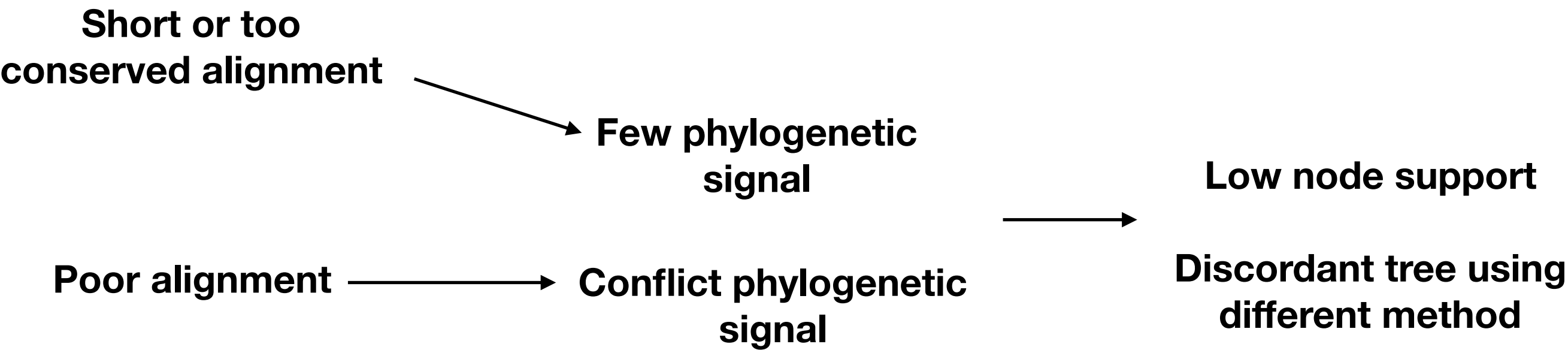
→ Tree reconstruction

`mafft_aln.pl`

Multiple sequence
alignment (MSA)

Align AA/DNA with
common ancestry

Why need to filter resulting alignment



Mis-alignment

Missing data

Paralogs

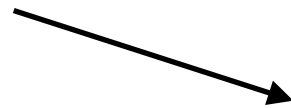
Saturated mutation

ATGCGTACGTT
ATGCGTATT--

ATGCGTACGTT
ATGCGTA--TT

Why need to filter resulting alignment

**Short or too
conserved alignment**



**Few phylogenetic
signal**



Low node support

Poor alignment



**Conflict phylogenetic
signal**

**Discordant tree using
different method**

Mis-alignment

Missing data

Paralogs

Saturated mutation

Filter poorly aligned regions

Poorly aligned coding regions

Remove sequences having long insertion
or deletion to reference



Remove sequences distant from reference



Remove sequences with low coverage



Realign

filter.pl

Filter poorly aligned regions

Sequences distant from reference

ref	CAGGACGTTTACTGAAGTTTTTCAGGAGAGCTTGAAGGTGTTTCAAGAGAAGAC
sample1	CTGGATGTCTGTTGAAGTTTTCTGGGGTGCTTGAAGACGTTTCAAGAGAGGAC
sample2	CTGGATGTCTGTTGAAGTTTTCTGGGGAGCTTGAAGACGTTTCAAGAGAGGAC
sample3	CTGGATGTCTGTTGAAGTTTTCTGGGGAGCTTGAAGATGTTTCAAGAGAGGAC
sample4	CTGGATGTCTGTTGAAGTTTTCTGGGGAGCTTGAAGATGTTTCAAGAGAGGAC
sample5	CTGGATGTCTGTTGAAGTTTTCTGGGGAGCTTGAAGATGTTTCAAGAGAGGAC
sample6	CTGGATGTCTGTTGAAGTTTTCTGGGGAGCTTGAAGATGTTTCAAGAGAGGAC

50 bp, 25 bp per step

Filter poorly aligned regions

Poorly aligned coding regions

**Remove sequences having long insertion
or deletion to reference**



Remove sequences distant from reference



Remove sequences with low coverage



Realign

Filter poorly aligned regions

Poorly aligned flanking regions

Remove unevenly enriched sequences from ends



Remove sequences having long and unique insertion



Remove too variable sequences



Remove short flanks and flanking sequences with low coverage



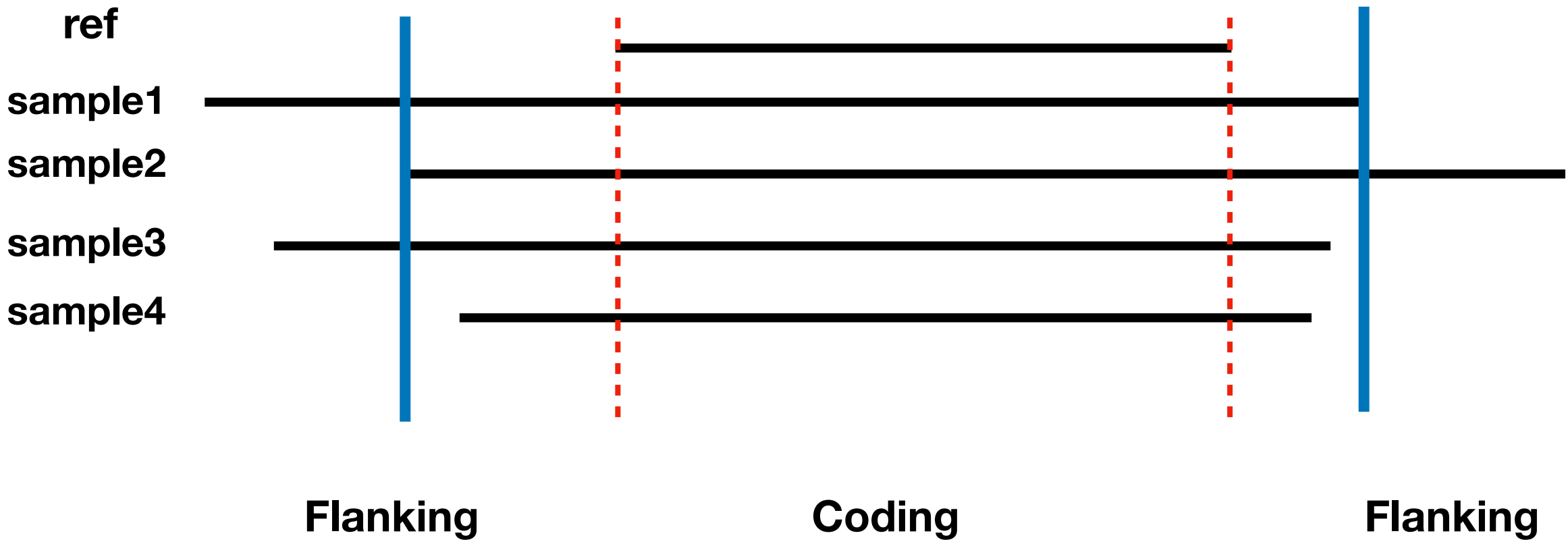
Realign

[flank_filter.pl](#)

Filter poorly aligned regions

Remove unevenly enriched sequences from ends

consecutive 5 columns
with 50% of residues



Filter poorly aligned regions

Poorly aligned flanking regions

Remove unevenly enriched sequences from ends



Remove sequences having long and unique insertion



Remove too variable sequences



Remove short flanks and flanking sequences with low coverage



Realign

Filter poorly aligned regions

Remove sequences having long and unique insertion



chimeric assembly

>= 10bp

Filter poorly aligned regions

Poorly aligned flanking regions

Remove unevenly enriched sequences from ends



Remove sequences having long and unique insertion



Remove too variable sequences



Remove short flanks and flanking sequences with low coverage

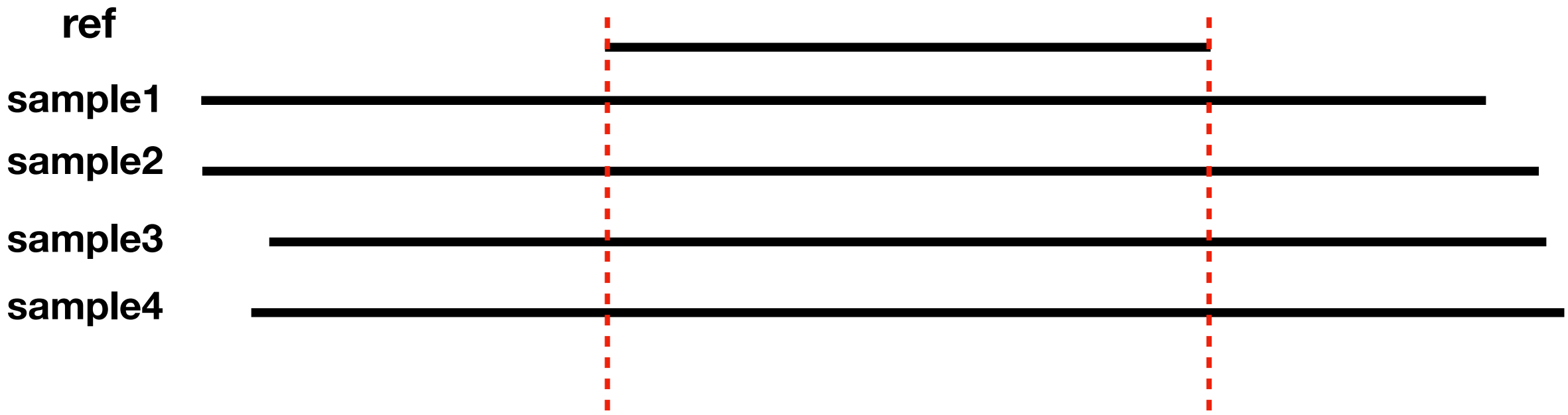


Realign

Filter poorly aligned regions

Remove too variable sequences

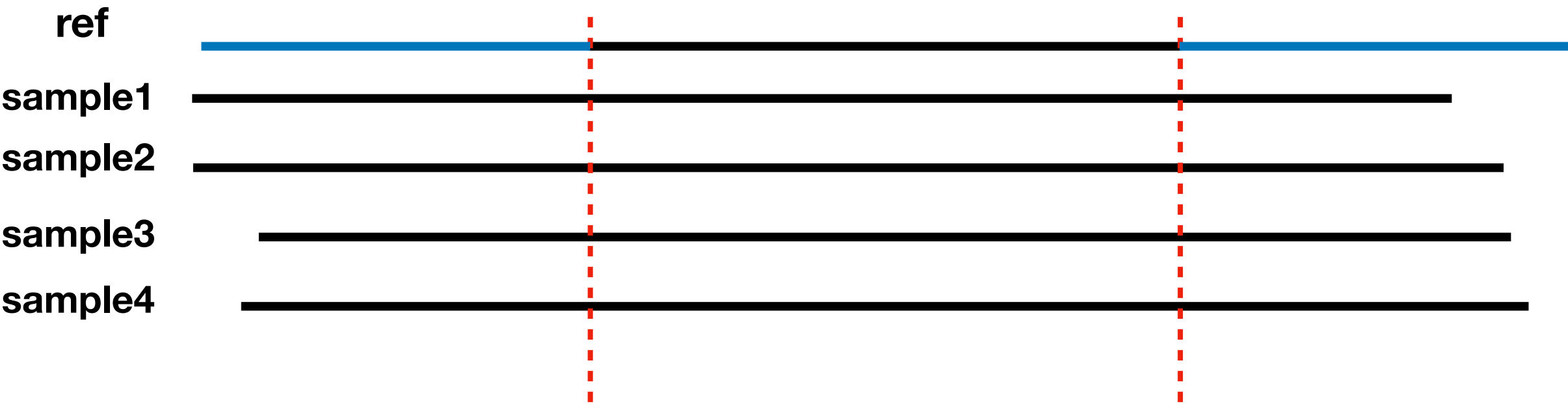
No reference in flanks



Filter poorly aligned regions

Remove too variable sequences

consensus reference



Remove sequences distant from consensus reference

Filter poorly aligned regions

Remove too variable sequences

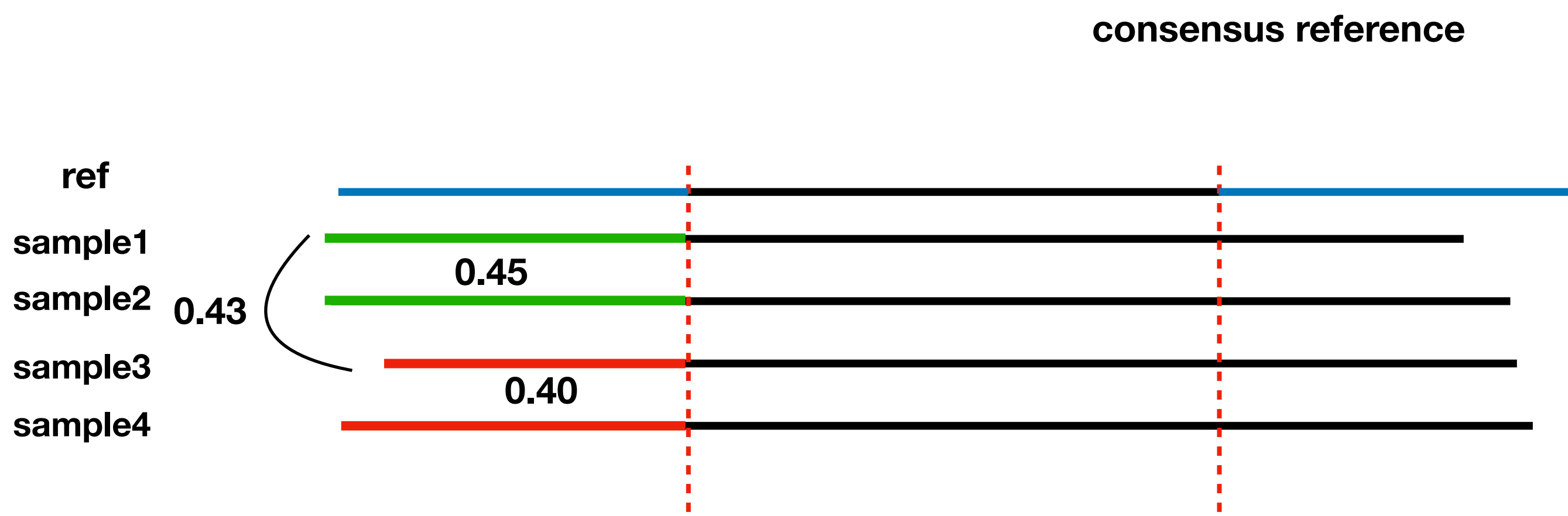
consensus reference



2 or more patterns of sequences existing in flanks

Filter poorly aligned regions

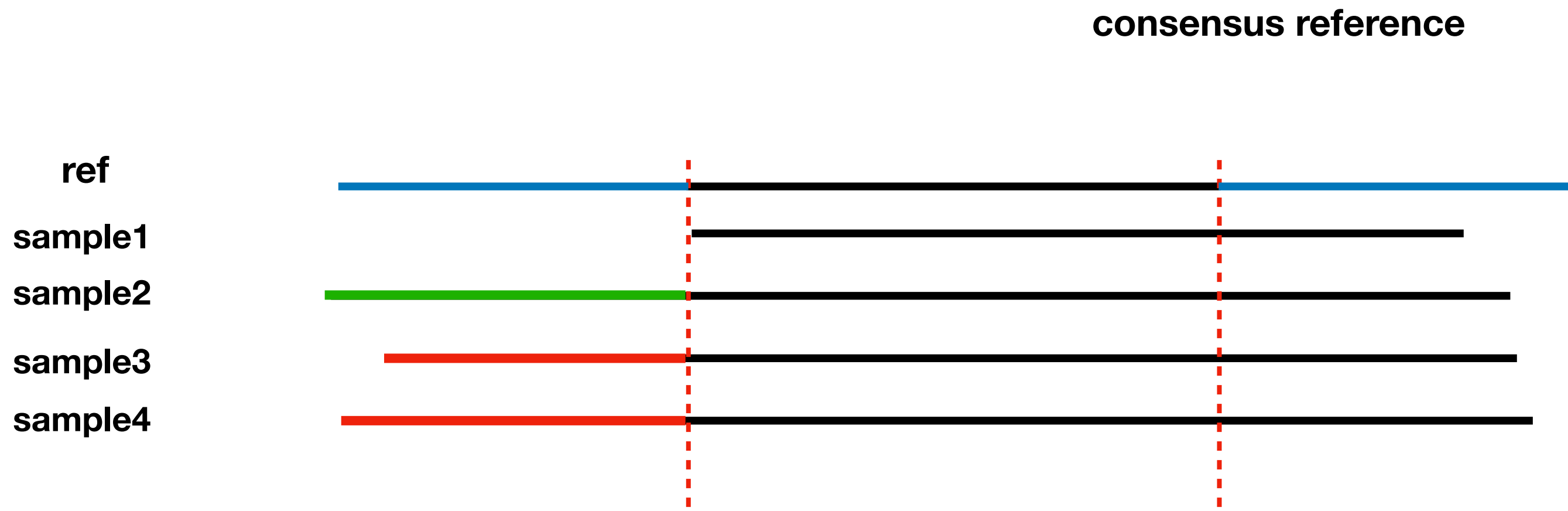
Remove too variable sequences



Find pair of distant sequences, then compute their distance to the rest of the sequences

Filter poorly aligned regions

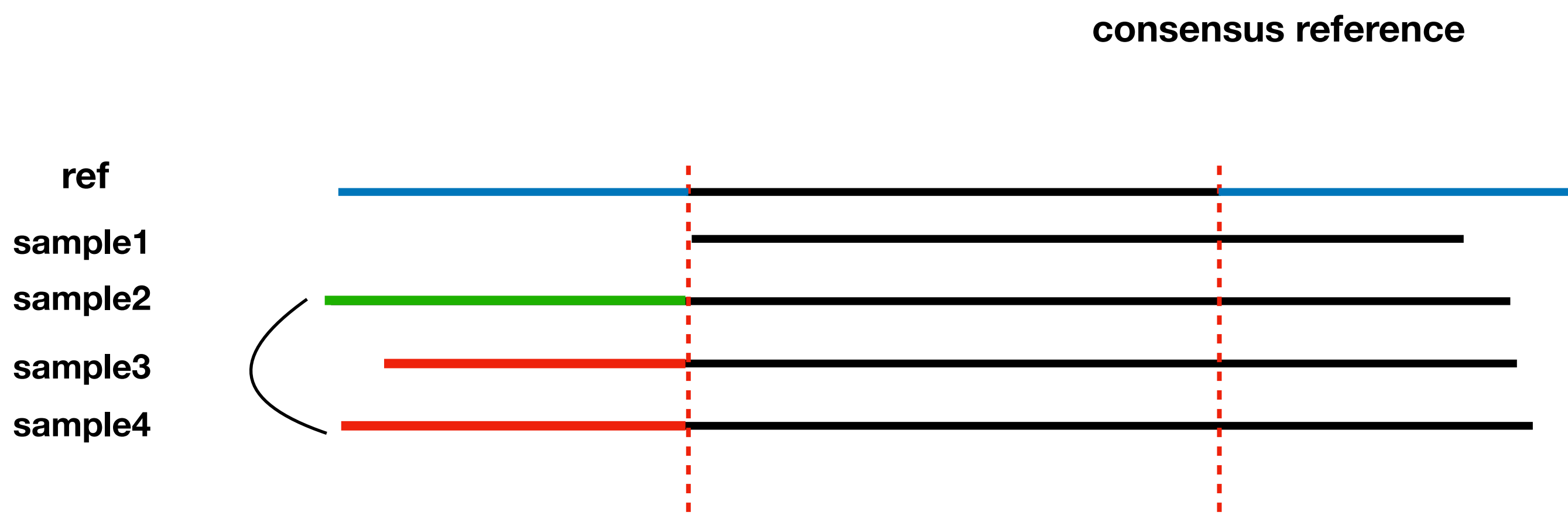
Remove too variable sequences



Remove sequences distant from rest of the sequences

Filter poorly aligned regions

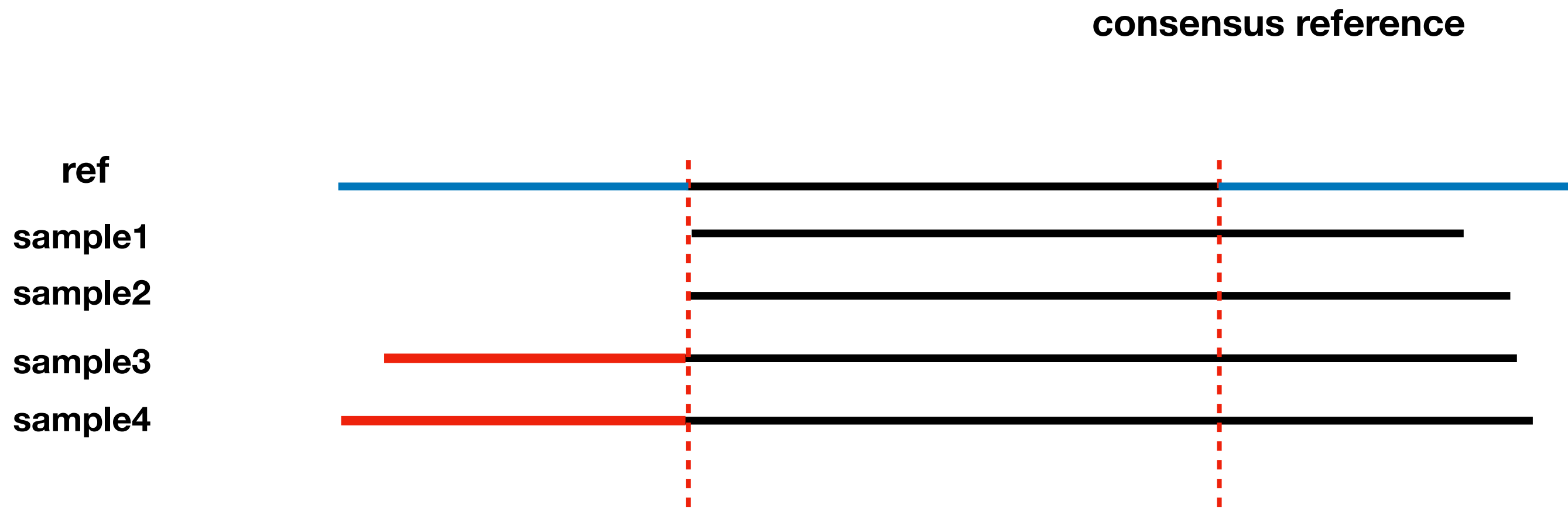
Remove too variable sequences



Remove sequences distant from rest of the sequences

Filter poorly aligned regions

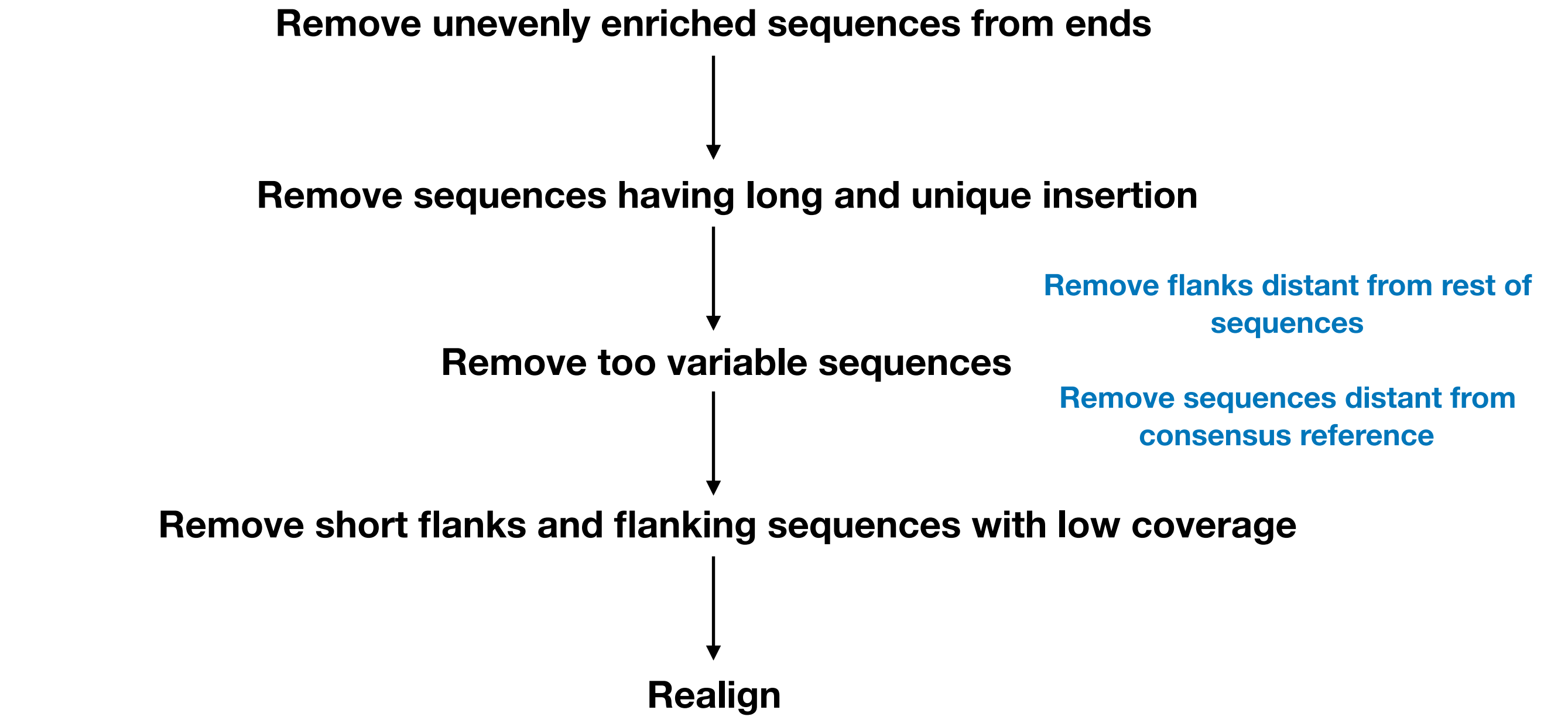
Remove too variable sequences



Remove sequences distant from rest of the sequences

Filter poorly aligned regions

Poorly aligned flanking regions




Filter poorly aligned regions

Poorly aligned flanking regions

Usage:

```
$ perl flank_filter.pl \
```

```
--flank assemble_result/f \
```



sequences with flanks

```
--nonflank_filtered filtered_nf \
```



filtered coding sequences

```
--flank_filtered Oreochromis_niloticus \
```



filtered sequences with flanks

```
--ref_taxa Oreochromis_niloticus
```



name of reference

Filter for other purpose

Filter out loci with pre-defined monophyletic group

Filter out loci follow the molecular clock hypotheses

Filter for other purpose

Filter out loci with pre-defined monophyletic group

```
sample1 sample2 sample3 sample4 sample5 sample6  
sample7 sample9  
sample8 sample10 sample3  
sample11 sample12
```

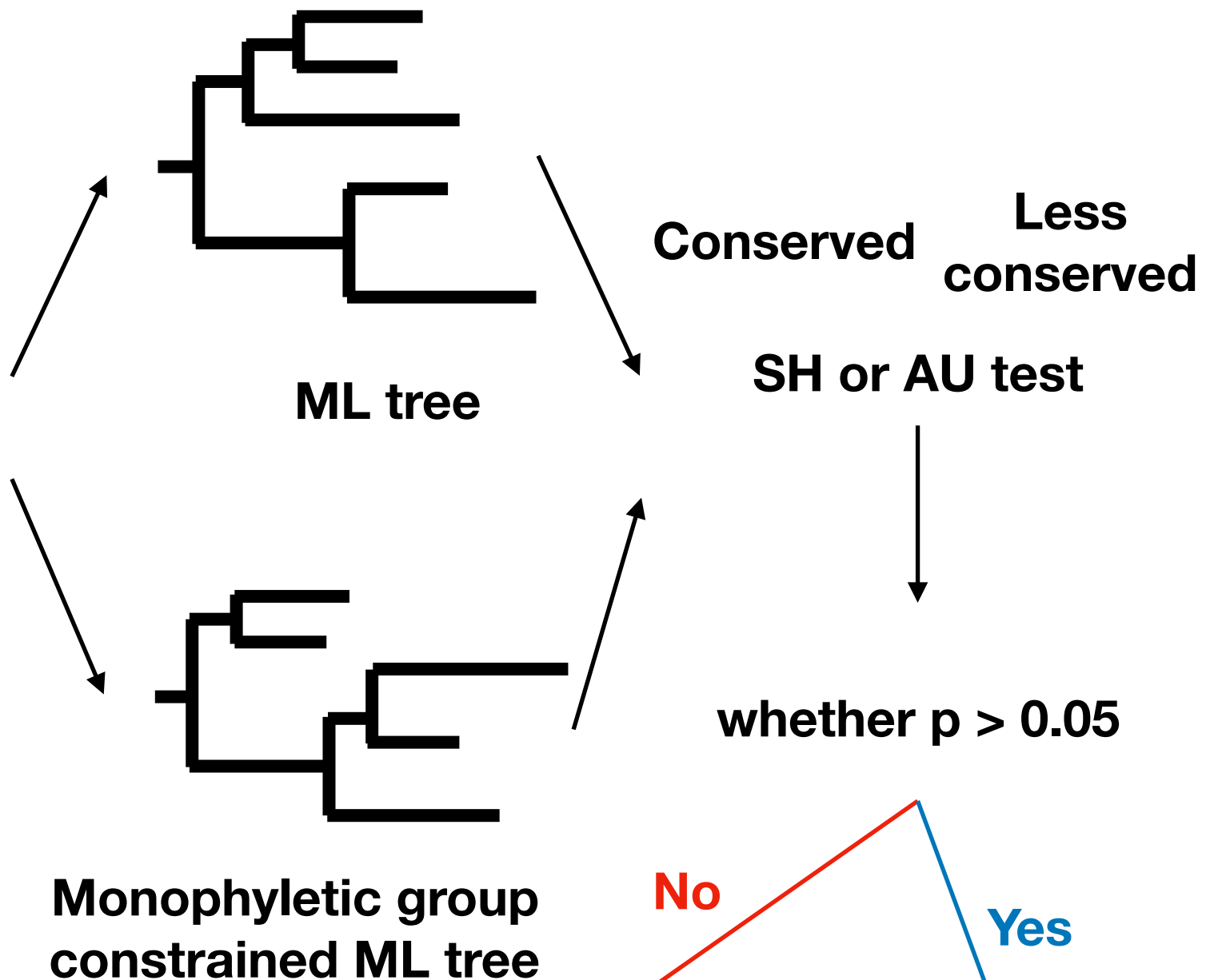
Define monophyletic group in a txt file, one group a line

At least **two sample in a group**

Filter for other purpose

SH and AU test

ATCGTAGGGCTGGCTAGTCGTAGCTA
ATCGTAGGGCTGGCGAGTCGT-GCTA
ATCGTAGGACTGGCTAGTCGTAGCTA
ATCGTACGGCTGGCTAGTCGTAGCTA
ATCGTAGGGCTGGCTAGTCGTAGCTA



use **construct_tree.pl** to build
constrained ML tree first

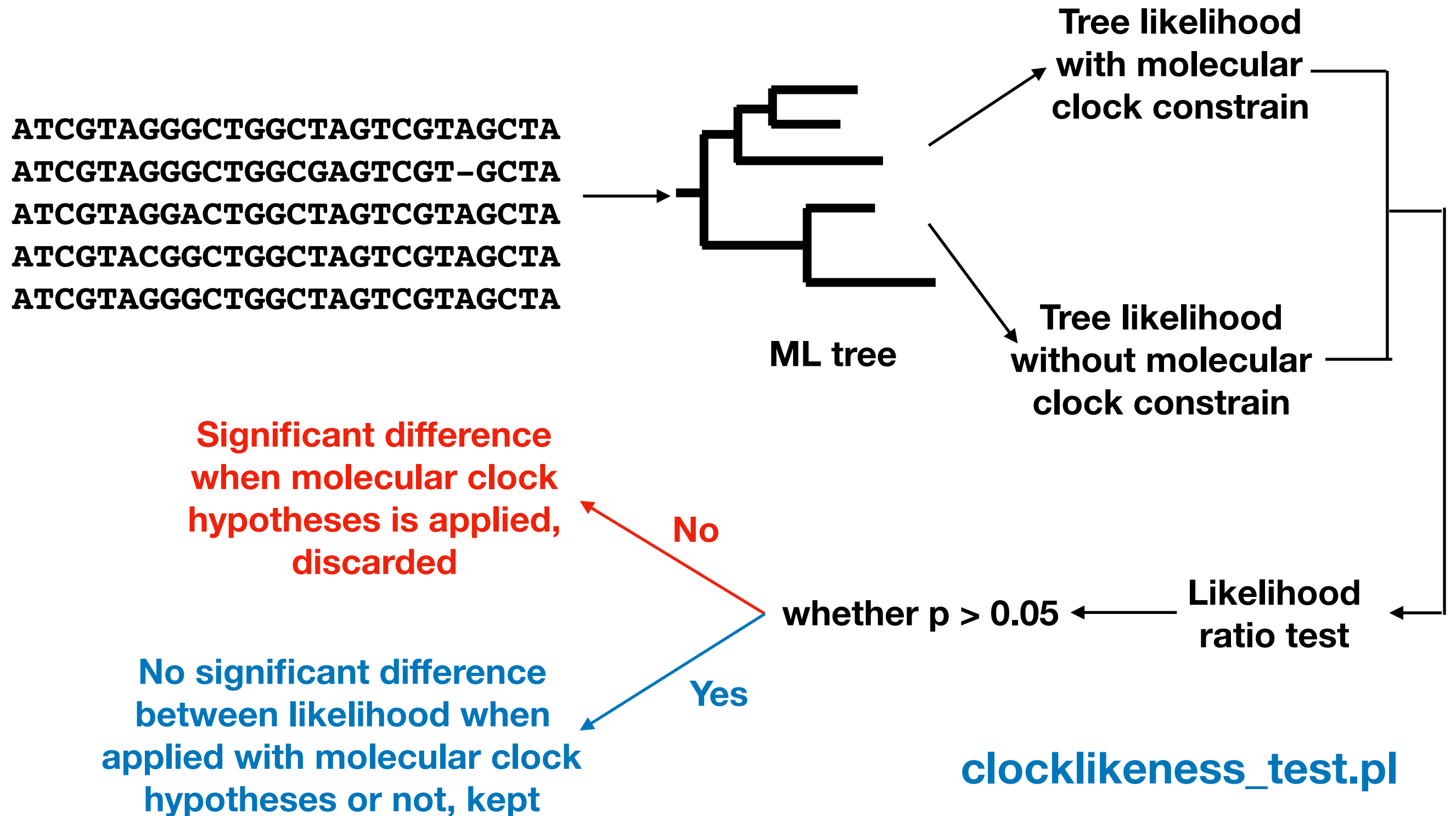
monophyly_test.pl

likelihood of two topologies with
given locus are significantly
different, so given locus do not
follow predefined group, discarded

likelihood difference of two
topologies with given locus
are not significant, so given
locus follows predefined
group, kept

Filter for other purpose

Filter out loci follow the molecular clock hypotheses



Detect cross-sample contamination

**Genetic distance of diverged taxa in most
loci cannot be very close**

[detect_contamination.pl](#)

Detect cross-sample contamination

```
sample1 sample2 sample3 sample4 sample5 sample6  
sample7 sample9  
sample8 sample10 sample3  
sample11 sample12
```

Define close related group in a txt file, one group a line

Permit **one sample in a group**

Detect cross-sample contamination

Closely related group1: Human Chimp Orangutan



**too close p-distance between
taxa (≤ 0.002)**

Closely related group2: Tilapia Zebra fish

Contamination rate (%) =

Potentially contaminated pair/Co-existence of this pair appeared in all loci*100

4434 loci in total

Potentially contamination between Human and Tilapia among all loci: 10 times

Co-existence of Human-Tilapia pair among all loci: 2000 times

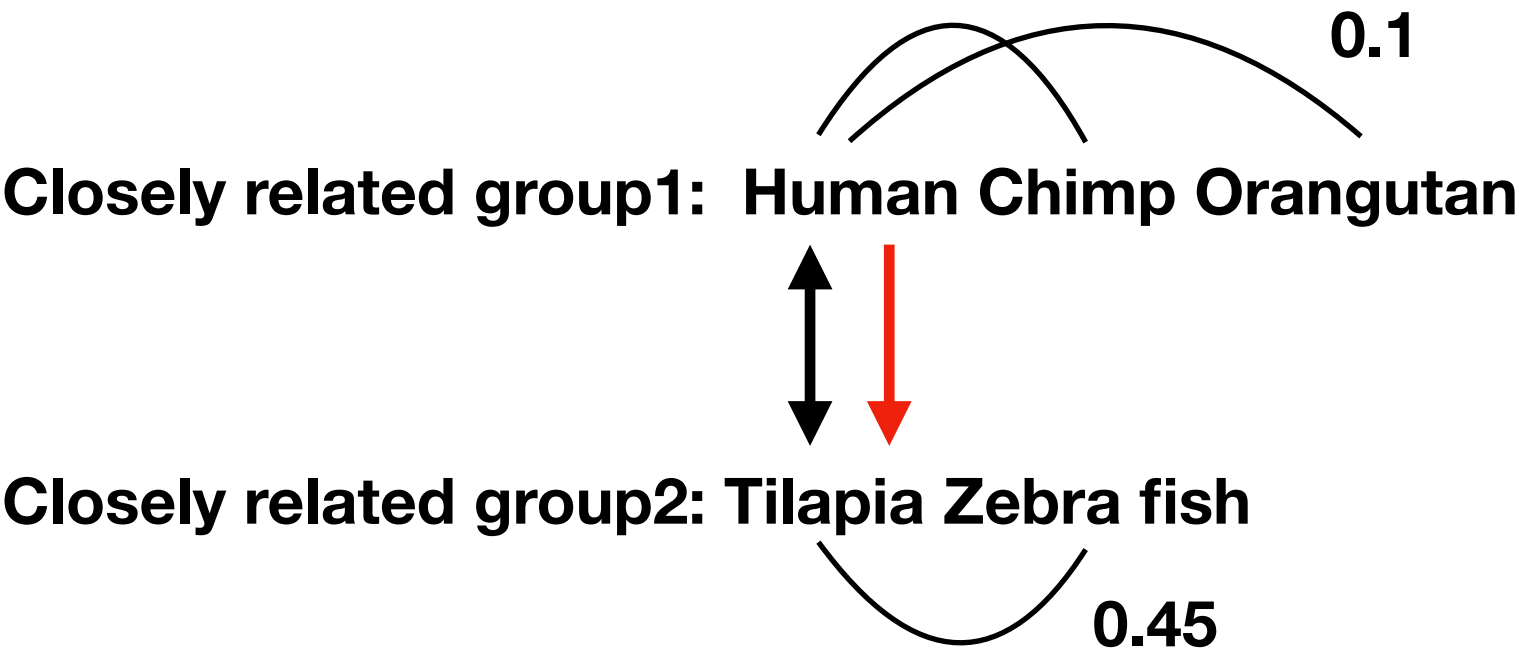
Contamination rate of Human-Tilapia (%) = $10/2000*100 = 0.5\%$

Default threshold:

$\geq 50\%$ Contamination rate

≥ 100 appearing time among all loci

Detect cross-sample contamination



Human <- Tilapia: 10
Human -> Tilapia: 200

Sample of Human contaminated Tilapia

Summary statistics

Summarized statistics for each **locus including:**

- (1) Average length of coding region**
- (2) Average length of flanking region**
- (3) Length of alignment**
- (4) Average GC content**
- (5) Percentage of Missing data**
- (6) Pairwise distance**

Summarized statistics for each **sample including:**

- (1) Average length of captured sequences**
- (2) Average GC content**
- (3) Number of captured loci**

Data filtering paradigm for coding region

ILS?
Excessively trimmed?
Too few data?

Contigs targeting
the same loci

Add sequences from existing
genomes if necessary

get_orthologues.pl

Filter data according to
completeness level pick_taxa.pl

taxa >= 80% & loci >= 500

No

remove loci <= 4 taxa

MSA mafft_aln.pl

Gene tree construct_tree.pl

Gene tree based Species
tree reconstruction

As expectation &
high node support

Yes



No

Filter or
trim loci

filter.pl,

detect_contamination.
pl, monophyly_test.pl,
clocklikeness_test.pl

Trouble
shooting

No enough data for
tree estimation

More data
required

Yes

remove
unqualified taxa

MAFFT mafft_aln.pl

ML Concatenated Gene tree based Species
tree reconstruction

concat_loci.pl

construct_tree.pl

As expectation &
Consistent

Yes

No

Trouble
shooting

Filter or
trim loci

filter.pl,
detect_contamination.
pl, monophyly_test.pl,
clocklikeness_test.pl

Conventional analysis paradigm

Interspecific analysis

Summarized statistics *statistics.pl*

Intraspecific analysis

